

The oxygen consumption rate of an inactive dry yeast (IDY) selected to protect wine from oxidation

Pere Pons¹, José M. Heras², Nathalie Sieczkowski², Joan Miquel Canals¹, Fernando Zamora¹ ¹ Department of Biochemistry and Biotechnology. Tarragona Oenology Faculty. Universidad Rovira i Virgili. C/ Marcel.lí Domingo 1.43002-Tarragona.

CHITOSAN

SPECIFIC INACTIVATED YEASTS

VINEYARD SOLUTIONS

² Lallemand Bio S.L. C/ Galileu 303. 1ª planta. 08028-Barcelona.

This article was published in InvestIgaclón y Ciencia, Jan / Feb 2019



Visionary biological solutions / www.lallemandwine.com

NUTRIENTS/PROTECTORS

ENZYMES

WINE YEASTS

WINE BACTERIA

Abstract

The oxygen consumption rate of a specific inactivated yeast (Pure-Lees™ Longevity, Lallemand) specially selected to protect wine from oxidation thanks to its high capacity for oxygen consumption was calculated using a non-destructive luminescence-based technique in a model solution, to which different concentrations of inactive dry yeast, sulphur dioxide or ascorbic acid were added. Results indicate that the specific inactivated yeast consumes oxygen at a similar rate to sulphur dioxide at the usual concentrations of use for both antioxidants.

Introduction

Ageing on lees, particularly white wines, is currently a widespread practice in winemaking [1]. The presence of lees in contact with the wine enriches the wine with polysaccharides and mannoproteins through the process of autolysis [2,3], which translates into increased mouth-feel [4,5] and, in the case of red wines, lower astringency [3,6]. Similarly, when it comes to sparkling wines, the presence of these macromolecules helps the incorporation of carbon dioxide gas and promotes foam stability [7,8]. Finally, the presence of mannoproteins aid the wine stability, helping to combat protein degradation [9] and prevent the appearance of tartaric acid salt crystals [10].

Another positive effect of the presence of lees on wine quality is their ability to lees consume oxygen [11,12], thereby protecting the wine from oxidation, both in terms of it's colour and it's aroma [4,13].

However ageing on lees, as well as being laborious, does entail certain risks such as the appearance of reduced aromas [4], increased production of histamines and other biogenic amines [14], or the development of problematic microorganisms such as *Brettanomyces/Dekkera* [14,15].

For this reason, the use of specific inactivated yeasts (SIY) has increased over the past few years as a source of mannoproteins and polysaccharides. SIYs enrich wines with these macromolecules without any of the aforementioned disadvantages [3,16]. More recently, it has been suggested that some SIYs may act to protect musts and wines from oxidation due to their high levels of glutathione [17], and also that some may directly consume oxygen [18].

Given the interest in looking for alternative antioxidants to sulphur dioxide for protecting wine from oxidation, the aim of this study is to evaluate the oxygen-consumption capacity of one SIY (Pure-Lees[™] Longevity. Lallemand), which has been specially selected for its high capacity for oxygen consumption, when compared to the two most commonly-used antioxidant additives, sulphur dioxide and ascorbic acid.

Materials and methods

The oxygen-consumption rate is calculated using a non-destructive luminescence-based technique, as described by Pascual et al, (2017) [19]. Figure 1 shows the experimental protocol used in this study.



Sensors (PreSens Precision Sensing GmbH, Regensburg, Germany) were attached to the insides of transparent bottles (0.75 L) to measure dissolved oxygen.



Figure 1. Experimental protocol to measure O₂ consumption in model wine by three different antioxidants.

These sensors were fixed to the centre of the inside of the bottle, which was filled with a model solution composed of 12% ethanol, 4 g tartaric acid/L, 3 mg/L of iron (III) and 0.5 mg/L of copper (II). The pH was adjusted to 3.5. This solution was saturated with oxygen by bubbling air through it for 10 minutes. To these bottles, 0, 50, 100 and 150 mg/l of SO₂ or ascorbic acid was added, or 200, 400 and 600 mg/L of the specific inactivated yeast (Pure-Lees[™] Longevity. Lallemand). The bottles were then hermetically sealed and kept at a constant temperature of 20°C. Oxygen concentration was measured for the first time one hour later, and was then measured periodically until oxygen consumption was complete.

Results

Figure 2 shows oxygen consumption by SO₂, ascorbic acid and the specific inactivated yeast over time. These graphs clearly show that all three antioxidants (SO₂, ascorbic acid and the specific inactivated yeast) do consume oxygen over time.



Figure 2. Oxygen consumption of three different antioxidants

It should be noted that ascorbic acid consumes oxygen much more rapidly than SO₂ and the specific inactivated yeast, as the time scale on this graph is hours, whereas the other antioxidants are measured in days.



Figure 3. Modelling of oxygen consumption by three different antioxidants

With the aim of modelling and parametrizing the kinetics of oxygen consumption, different models of adjustment were tried. Representation of the inverse oxygen consumed compared to inverse time was the model that provided the best linear adjustment. **Figure 3** demonstrates the results obtained. It can be seen that excellent linear regression coefficients were obtained, which confirms that this mathematical model works well. Using this model, the following equation can be established.

$1 / [O_2] = a / t + B.$

This equation describes the relationship between oxygen consumed and the time function which allows for, after carrying out the first derivative and considering the baseline (time zero), the calculation of the oxygen consumption rate (OCR) which corresponds to the inverse of the equation slope. These calculations can be seen in **Figure 4**.



Figure 4. Calculation of the baseline (time zero) oxygen consumption rate (bOCR)

With the slopes obtained through applying this model of adjustment it is possible to calculate the relationship between concentration of SO₂, ascorbic acid or specific inactivated yeast and the baseline oxygen consumption rate (bOCR). **Figure 5** illustrates the result.



Figure 5. Oxygen consumption rates (bOCR0) for SO₂, ascorbic acid and specic inactivated yeast

The results demonstrate that there is a good linear relationship between oxygen consumption and the concentration of the three antioxidants. The slopes of the regression lines correspond to the oxygen consumption rates as reflected in **Table 1**. In this table, it can be seen that ascorbic acid has a much higher oxygen consumption rate than sulphur dioxide or specific inactivated yeast (around 300 times faster). However, it should be taken into account that ascorbic acid generates hydrogen peroxide, and its use without the addition of SO₂ can generate greater oxidation after a certain length of time [20]. However, the specific inactivated yeast Pure-Lees[™] Longevity consumes oxygen when applied at 40g/hL at a rate of 0.29 mg of oxygen/day. This value is very similar to sulphur dioxide at a dose of 20 mg/L (0.24 mg of oxygen/day), meaning that its efficacy can be considered to be equivalent to that of SO₂ at its usual levels of use.

Antioxidant –		Oxygen consumption rate (OCR) mg O ₂ /day		
2.976	148.8	297.6	446.4	
SO2	mg O ₂ /mg SO ₂ .day	[SO ₂] = 20 mg/L	[SO ₂] = 40 mg/L	[SO ₂] = 60 mg/L
	0.0119	0.24	0.48	0.71
Pure-Lees™ Longevity	mg O ₂ /mg SIY.day	[SIY] = 20 mg/hL	[SIY] = 40 mg/hL	[SIY] = 60 mg/hL
	0.0072	0.14	0.29	0.43

Table1. Oxygen consumption rates (OCR) for SO₂, ascorbic acid and specific inactivated yeast



Conclusion

The results show that the specific inactivated yeast Pure-LeesTM Longevity consumes oxygen in a model medium at a similar rate to sulphur dioxide, and therefore could be a good alternative to SO_2 and a means to reduce the levels of SO_2 used during winemaking, while still protecting wine from oxidation.

Acknowledgements

This study was funded by CDTI (CIEN Programme) "new strategies in the wine sector for sustainability and increased competitiveness on the international market (VINYSOST 2014)."

Bibliography

- 1. Zamora F. (2002) La crianza del vino tinto sobre lías; Una nueva tendencia. *Enólogos*, 19, 24-29.
- 2. González, E., Urtasun, A., Gil, M., Kontoudakis, K., Esteruelas, M., Fort, F., Canals, J.M., Zamora, F. (2013) Effect of yeast strain and supplementation with inactive yeast during alcoholic fermentation on wine polysaccha- rides. *Am. J. Enol. Vitic.*, 64, 268-273.
- 3. González-Royo, E., Esteruelas, M., Kontoudakis, N., Fort, F., Canals, J.M., Zamora, F. (2017) The effect of supplementation with three commercial inactive dry yeasts on the color, phenolic compounds, polysaccharides and astringency of a model wine solution and red wine. *J. Sci. Food Agric.*, 97, 172–181.
- 4. Fornairon-Bonnefond, C., Camarasa, C., Moutounet, M., Salmón, J.M. (2002) New trends on yeast autolysis and wine ageing on lees: a bibliographical review. *J. Int. Vigne Vin* 36, 49-69.
- Del Barrio-Galán, R., Pérez-Magariño, S., Ortega-Heras, M., Williams, P., Doco, T. (2011) Effect of aging on lees and of three different dry yeast derivative products on verdejo white wine composition and sensorial characteristics. *J. Agric. Food Chem.*, 59,12433–12442.
- Quijada-Morín, N., Williams, P., Rivas-Gonzalo, J.C., Doco, T., Escribano-Bailón, M.T. (2014) Polyphenolic, polysaccharide and oligosaccharide composition of tempranillo red wines and their relationship with the perceived astringency. *Food Chem.*, 154. 44-5.
- Esteruelas, M., González-Royo, E., Kontoudakis, N., Orte, A., Cantos, A., Canals, J.M., Zamora, F. (2015) Influence of Grape Maturity on the Foaming Properties of Base Wines and Aparkling Wines (cava). J. Sci. Food Agric., 95, 2071-2080.
- 8. González-Royo, E., Pascual, O., Kontoudakis, N., Este- ruelas, M., Esteve-Zarzoso, B., Mas, A., Canals, J.M., Zamora, F. (2015) Oenological consequences of sequential inoculation with non-*Saccharomyces* yeasts (*Torulaspora delbrueckii* or *Metschnikowia pulcherrima*) and *Saccharomyces cerevisiae* in base wine for sparkling wine production. *Eur. Food Res. Technol.*, 240, 999-1012.
- 9. Lubbers, S., Leger, B., Charpentier, C., Feuillat, M. (1993) Effet des colloïdes-protecteurs d'extraits de parois de levures sur la stabilité tartrique d'un vin modè- le. *J. Int. Sci. Vigne Vin* 27,13-22.





- 11. Fornairon, c., Mazauric, J.P., Salmon, J.M., Moutounet, M. (1999) Observations sur la consommation de l'oxygène pen- dant l'élevage des vins sur lies. *J. Int. Vigne Vin*, 33, 79-86.
- Salmon, J.M., Mazauric, J.P., Fornairon, C. Y Moutou- net, M.(2001) L'enjeu œnologique de l'élevage sur lie des vins rouges. I – Les lies de levure et la consommation d'oxygène. en "connaissances actuelles & avenir de l'élevage en barriques. *Burdeos*, pp 39-42.
- Salmon, J.M., Fornairon-Bonnefond, C., Mazauric, J.P., Moutounet, M. (2000) Oxygen consumption by wine lees: impact on lees integrity during wine ageing. *Food Chem.*, 71, 519-528.
- 14. Pérez-Serradilla, J.A., Luquede Castro, M.D. (2008) Role of lees in wine production: a review. *Food Chem.*, 111, 447-456.
- 15. Oelofse, A., Pretorius, I.S., du Toit, M. (2008) Significance of *Brettanomyces* and *Dekkera* during Winemaking: a synoptic Review. s. afr. *J. Enol. Vitic.*, 29, 128-144.
- Pozo-Bayón, M.A., Andujar-Ortiz, I., Alcaide-Hidalgo, J.M., Martín-Álvarez, P.J., Moreno-arribas, M.v. (2009) characterization of commercial inactive dry yeast preparations for enological use based on their ability to release soluble compounds and their behavior toward aroma compounds in model wines. *J. Agric. Food Chem.*, 57, 10784-10792.
- Andújar-Ortiz, I., Chaya, C., Martín-Álvarez, P.J., Moreno-Arribas, M.V., Pozo-Bayón, M.A. (2014) Impact of using new commercial glutathione enriched inactive dry yeast oenological preparations on the aroma and sensory properties of wines. *Int. J. Food Prop.*, 17, 987-1001.
- 18. Pons, P., Heras, J.M., Sieczkowski, N., Canals, J.M., Zamora, F. (2018) Determinación de la tasa de consumo de oxígeno por parte de una levadura seca inactivada (LsI); comparación con el ácido ascórbico y con el dióxi- do de azufre. Comunicación al XIV Congreso Nacional de Investigación Enológica. GIENOL 2018.
- Pascual, O., Vignault, A., Gombau, J., Navarro, M., Gómez-Alonso, S., García-Romero, E., Canals, J.M., Her- mosín-Gutíerrez, I., Teissedre, P.L., Zamora, F. (2017) Oxygen consumption rates by different oenological tannins in a model wine solution. *Food Chem.*, 234, 26-32.
- 20. Peng, Z., Duncan, B., Pocock, K.F., Sefton, M.A. (1998) The effect of ascorbic acid on oxidative browning of white wines and model wines. *Aust. J. Grape Wine Res.*, 4, 127-135.